**ORIGINAL PAPER** 



# Mathematical analysis and drug exposure evaluation of pharmacokinetic models with endogenous production and simultaneous first-order and Michaelis–Menten elimination: the case of single dose

Xiaotian Wu<sup>1,2</sup> · Fahima Nekka<sup>2,3</sup> · Jun Li<sup>2,3</sup>

Received: 8 February 2018 / Accepted: 30 June 2018 / Published online: 9 July 2018 © Springer Science+Business Media, LLC, part of Springer Nature 2018

#### Abstract

Drugs with an additional endogenous source often exhibit simultaneous first-order and Michaelis–Menten elimination and are becoming quite common in pharmacokinetic modeling. In this paper, we investigate the case of single dose intravenous bolus administration for the one-compartment model. Relying on a formerly introduced transcendent function, we were able to analytically express the concentration time course of this model and provide the pharmacokinetic interpretation of its components. Using the concept of the corrected concentration, the mathematical expressions for the partial and total areas under the concentration time curve (AUC) were also given. The impact on the corrected concentration and AUC is discussed as well as the relative contribution of the exogenous part in presence of endogenous production. The present findings theoretically elucidate several pharmacokinetic issues for the considered drug compounds and provide guidance for the rational estimation of their pharmacokinetic parameters.

**Keywords** Pharmacokinetic model  $\cdot X$  function  $\cdot$  Endogenous production  $\cdot$  Simultaneous first-order and Michaelis–Menten elimination  $\cdot$  Area under the concentration time curve (AUC)

### Introduction

In clinical pharmacology, quantitative pharmacokinetic modeling has been proved to effectively link the administered drug amount and the induced therapeutic outcome by describing the complete concentration time course for the desired and/or undesired effect [1, 2]. With the increasing complexity involved in these models, their quantitative

**Electronic supplementary material** The online version of this article (doi:https://doi.org/10.1007/s10928-018-9599-4) contains supplementary material, which is available to authorized users.

Fahima Nekka fahima.nekka@umontreal.ca

- <sup>1</sup> Department of Mathematics, Shanghai Maritime University, Shanghai 201306, People's Republic of China
- <sup>2</sup> Faculté de pharmacie, Université de Montréal, Montréal, QC H3C 3J7, Canada
- <sup>3</sup> Centre de recherches mathématiques, Université de Montréal, Montréal, QC H3C 3J7, Canada

analysis and interpretation become determinant for the prediction quality of direct observations and other pharmacological properties. Historically, linear pharmacokinetic compartment models have greatly contributed to the explanation and prediction of many pharmacological phenomena. However, many of new drug compounds, such as hormones or monoclonal antibodies, can have more complex kinetics [3-6]. A typical example is the recombinant human granulocyte colony-stimulating factor (rhG-CSF), which is molecularly similar to endogenous produced granulocyte colony-stimulating factor (G-CSF) and generally used as a stimulant agent to mitigate the toxic effect of chemical drugs on white blood cells during chemotherapy. G-CSF functions by promoting the generation, differentiation and maturation of neutrophils, a major part of white blood cells that play a crucial role in the immune system [3, 4, 6]. The elimination of G-CSF not only undergoes the traditional renal pathway, generally of linear type, but also combines a saturate internalization process, in which the G-CSF molecules bind to neutrophil receptors and are transformed into substances readily to be eliminated. Other blood stimulants, such as erythropoietin (EPO) and thrombopoietin (TPO) show similar PK properties [7–9].

In order to predict plasma concentration time courses of these drugs, compartment models with nonlinear elimination is a reasonable choice. The general approach is to model the saturate drug elimination with Michaelis-Menten kinetics [10–12]. However, refined models using parallel elimination mechanisms, with a first-order kinetics characterizing the renal elimination pathway and a Michaelis-Menten kinetics for the saturate metabolism pathway, were proposed as well [6, 9, 13]. Many efforts are solely based on numerical solutions, and are limited to pointing out the existence of an implicit function for the solution [14, 15]. However, it would be helpful to concretely find a closed form solution, which allows to explore the underlying mechanisms and easily extract various relevant pharmacological properties. It has become a challenge to search for analytical solutions of these complex models. For instance, in the case of compartment models with Michaelis-Menten elimination alone, Lambert W function proved to be the sought-for element able to express their solutions under a closed form [16, 17]. Specifically, the time course of drug plasma concentration  $C_m(t)$  after a single intravenous (IV) bolus dose administration D can be expressed as

$$C_m(t) = K_m \cdot W\left(\frac{D}{K_m V_d} \exp\left(\frac{D - V_{max}t}{V_d K_m}\right)\right),\tag{1}$$

where "W" represents Lambert W function [18];  $V_{max}$  is the maximum velocity of Michaelis–Menten kinetics in unit of amount/time, and  $K_m$ , known as the Michaelis–Menten constant, is the concentration value at which the rate of change of Michaelis–Menten kinetics reaches half of  $V_{max}$ ;  $V_d$  is the body's apparent volume of distribution. Recently, motivated by Lambert W function, we introduced the X function and obtained the corresponding closed form solution of C(t) for drugs modeled with a one-compartment structure and having simultaneous first-order ( $k_{el}$ ) and Michaelis–Menten elimination ( $K_m$  and  $V_{max}$ ) [19]. For the case of a single dose D, C(t) is therefore expressed as

$$C(t) = C_{\beta} \cdot X\left(\left(\frac{D}{C_{\beta}V_d}\right)^{p_1} \left(\frac{D}{C_{\beta}V_d} + 1\right)^{p_2} \exp(-t), p_1, p_2\right),$$
(2)

where

$$p_{1} = \frac{K_{m}V_{d}}{k_{el}V_{d}K_{m} + V_{max}}, \quad p_{2} = \frac{V_{max}}{k_{el}(k_{el}V_{d}K_{m} + V_{max})},$$
$$C_{\beta} = K_{m} + \frac{V_{max}}{k_{el}V_{d}}.$$
(3)

Note that the symbol  $C_{\beta}$  used here replaces  $\beta$  that was formally used in [19] to make the content easier to understand.

Apart from their simultaneous linear and saturate elimination properties, compounds such as hormones or monoclonal antibodies can have more complex properties due to their endogenous production. While endogenous production has been accounted for in compartment PK models, it only remained at the level of parameters fitting of plasma concentration data [3, 20]. The systematic analysis of these models is still absent, either for closed form solutions, or for model-based estimation of PK parameters, particularly for the area under concentration time curve (AUC). Even though in the linear case, where several algebraic formulas have been established and used to estimate such parameters in practice, the rationale behind these estimations is still lacking when endogenous production is present. It is therefore important to understand how the endogenous production can impact the disposition of these molecules. Consequently, the mathematical validity of current AUC estimation should be revisited. This will be the focus of the present work.

The paper is organized as follows: the relationship between *Lambert W* and *X* functions is clarified in the next section, followed by the closed form solution for the model of single IV bolus administration and the PK interpretation of newly introduced entities. The analytical formulas for the estimation of AUC in the case of constant endogenous production using the corrected concentration and the impact of endogenous production on the corrected concentration and AUC are then studied. Finally, the limits and perspectives of the current PK models, either from a pharmacological or a bio-mathematical standpoint are discussed in the last section.

# Lambert W and X functions

*Lambert W* function is a transcendental function widely used for the closed form solutions of various differential equations describing exponential phenomena, particularly for delay differential equations [21, 22]. In pharmacokinetics, it has been used to express the closed form solutions of drug plasma concentration time courses when only Michaelis–Menten elimination is involved [16, 17]. In order to analytically express solutions of one-compartment PK models of simultaneous first-order and Michaelis–Menten elimination, we have introduced the *X* function as a natural extension of *Lambert W* function [19]. As shown below, we can see how these two functions are related.

**Definition 1** [18] The *Lambert W* function is the multivalued inverse of the function  $f(z) = z \exp(z)$ , i.e.,

$$W(z)\exp(W(z)) = z \tag{4}$$

where z is a complex number.

**Definition 2** [19] The X function is the multivalued inverse of the function  $f(z) = z^p(z+1)^q$ ,  $p, q \in \mathbb{R}^+$ , i.e.,

$$(X(z, p, q))^{p}(X(z, p, q) + 1)^{q} = z$$
(5)

where z is a complex number.

The PK model having simultaneous first-order and Michaelis–Menten elimination is an extension of the model involving only a Michaelis–Menten elimination. Hence the X function can be considered an extension to the *Lambert* W function. However, this extension is neither simple nor direct. In fact, instead of being a particular case of the X function, the *Lambert* W function can only be obtained through a limit process of the former.

From Eq. (2), we have

$$\left(\frac{C(t)}{C_{\beta}}\right)^{p_1} \left(\frac{C(t)}{C_{\beta}} + 1\right)^{p_2} = \left(\frac{D}{C_{\beta}V_d}\right)^{p_1} \left(\frac{D}{C_{\beta}V_d} + 1\right)^{p_2} \exp(-t)$$

which can be further rearranged into

$$(C(t))^{p_1} \left(\frac{C(t)}{C_{\beta}} + 1\right)^{p_2} = \left(\frac{D}{V_d}\right)^{p_1} \left(\frac{D}{C_{\beta}V_d} + 1\right)^{p_2} \exp(-t),$$
(6)

where  $p_1$ ,  $p_2$  and  $C_\beta$  are given in Eq. (3). Using the dependence of the models on these parameters, we have

$$\lim_{k_{el}\to 0} C(t) = C_m(t).$$

When taking  $k_{el} \rightarrow 0$ , it can be proved that

$$\lim_{k_{el}\to 0} p_1 = \frac{K_m V_d}{V_{max}}, \quad \lim_{k_{el}\to 0} \frac{p_2 C(t)}{C_\beta} = \frac{V_d C_m(t)}{V_{max}}$$
  
and 
$$\lim_{k_{el}\to 0} \left(\frac{C(t)}{C_\beta} + 1\right)^{\frac{C_\beta}{C(t)}} = e$$

where e is the Euler's number.

Using the above relationships, we have

$$\lim_{k_{el}\to 0} \left(\frac{C(t)}{C_{\beta}} + 1\right)^{p_2} = \lim_{k_{el}\to 0} \left[ \left(\frac{C(t)}{C_{\beta}} + 1\right)^{\frac{C_{\beta}}{C(t)}} \right]^{\frac{p_2C(t)}{C_{\beta}}}$$
$$= \lim_{k_{el}\to 0} \left[ \left(\frac{C(t)}{C_{\beta}} + 1\right)^{\frac{C_{\beta}}{C(t)}} \right]^{\frac{lim}{k_{el}\to 0}\frac{p_2C(t)}{C_{\beta}}}$$
$$= \exp\left(\frac{V_d C_m(t)}{V_{max}}\right).$$

Moreover, we have

$$\lim_{k_{el}\to 0} \left(\frac{D}{C_{\beta}V_d} + 1\right)^{p_2} = \exp\left(\frac{D}{V_{max}}\right)$$

For both sides of Eq. (6), we take the limit as  $k_{el} \rightarrow 0$ , then

$$(C_m(t))^{\frac{V_d K_m}{V_{max}}} \exp\left(\frac{V_d C_m(t)}{V_{max}}\right) = \left(\frac{D}{V_d}\right)^{\frac{1}{M_{max}}} \exp\left(\frac{D}{V_{max}}\right) \exp(-t).$$

Simplifying the above equation, we have

$$\frac{C_m(t)}{K_m} \exp\left(\frac{C_m(t)}{K_m}\right) = \frac{D}{V_d K_m} \exp\left(\frac{D - V_{max}t}{V_d K_m}\right).$$
(7)

If we use *Lambert W* function to express the solution of Eq. (7), it will correspond to the one given by Eq. (1).

Following the definitions of *Lambert W* and *X* functions, the relation between these two functions can be summarized as:

**Theorem 1** Write  $z = \frac{D}{V_d K_m} \exp\left(\frac{D - V_{max} t}{V_d K_m}\right)$ , then Lambert W function is a limit of X functions as  $k_{el} \rightarrow 0$ , which can be expressed in the following limit form:

$$\lim_{k_{cl}\to 0} \frac{C_{\beta}}{K_{m}} \cdot X\left(\left(\frac{D}{C_{\beta}V_{d}}\right)^{p_{1}} \left(\frac{D}{C_{\beta}V_{d}}+1\right)^{p_{2}}\right)$$
$$\exp\left(-\frac{D}{V_{max}}\right) \left(\frac{K_{m}V_{d}}{D}z\right)^{\frac{V_{d}K_{m}}{V_{max}}}, p_{1}, p_{2}\right) = W(z).$$

**Remark 1**  $K_m W\left(\frac{D}{K_m V_d} \exp\left(\frac{D - V_{max}t}{V_d K_m}\right)\right)$  is the solution

of the PK model with the Michaelis–Menten elimination alone (Eq. 1), and  $C_{\beta}X\left(\left(\frac{D}{C_{\beta}V_d}\right)^{p_1}\left(\frac{D}{C_{\beta}V_d}+1\right)^{p_2}\exp(-t), p_1, p_2\right)$  is the solution of the model with simultaneous first-order and Michaelis–Menten elimination (Eq. 2). The former is the limit of the latter when the linear elimination tends to fade.

As for the case of *Lambert W* function, the X function has multiple real branches. However there is only one real branch in the first quadrant, which we need to express the analytical solutions of PK models. In the rest of the paper, we will use the symbol X to denote this unique real branch. A brief discussion of this topic is given in the Appendix 4.

It is practical to have the X function implemented into mathematical software such as *Matlab*, which is what we have done here to compute the drug plasma concentration of the discussed PK model.

# Pharmacokinetic model of endogenous production and simultaneous first-order and Michaelis–Menten elimination

The X function was introduced to express the closed form solution of a PK model with simultaneous first-order and Michaelis–Menten elimination in a previous work [19]. For the more general PK model considered in the current paper, we will show how the X function can serve its closed form solution.

### The description of the PK model

As mentioned in the introduction, it is not rare that drug substances (given exogenously) are also endogenously produced, and eliminated through parallel pathways [3, 23, 24]. Indeed, their elimination can involve a first-order process generally through kidneys, in a proportional way to the drug plasma concentration, accompanied by a non-linear elimination of Michaelis–Menten kinetics, likely attributed to drug-mediated metabolism or internalization. It is usually assumed that endogenous production occurs at a constant rate, denoted by  $r_{prod}$ , if circadian effect can be ignored [3]. The following differential equation is used to describe the PK model considered here:

$$\frac{d}{dt}C(t) = r_{prod} - k_{el}C(t) - \frac{1}{V_d}\frac{V_{max}C(t)}{K_m + C(t)}, \quad t > 0, \qquad (8)$$

with initial conditions

$$C(0^{+}) = C_{hs} + D/V_d^{def} C_0, \ at \ t = 0^{+},$$
(9)

where  $k_{el}$ ,  $V_{max}$ ,  $K_m$ ,  $V_d$ , D are as previously defined, and  $C_{hs}$  having the form

$$C_{hs} = \frac{1}{2} \left( \frac{r_{prod}}{k_{el}} - C_{\beta} + \sqrt{\left(\frac{r_{prod}}{k_{el}} - C_{\beta}\right)^2 + 4\frac{r_{prod}}{k_{el}}K_m} \right)$$
(10)

is the baseline concentration calculated from the system at homeostasis. The well-posedness of the model (8) is provided in Appendix 1.

It is worth noting that the baseline concentration can be estimated prior to drug administration [25–27]. We also have to mention that if  $r_{prod} = 0$ , the current model goes back to the model with simultaneous first-order and Michaelis–Menten elimination studied in [19]. Moreover, without loss of generality and for the rest of the paper, the concentration value immediately after dose administration is referred to as the concentration at time zero (Eq. 9).

#### Springer

#### Closed form solution of C(t)

The closed form solution of the concentration time course of the considered model (Eqs. 8–9) can be expressed as follows.

**Theorem 2** For an intravenous bolus dose D, the closed form solution of Eqs. (8)–(9) is

$$C(t) = C_{hs} + \left(C_{hs} + C_{\beta}^{en}\right) \cdot X\left(\left(\frac{D/V_d}{C_{hs} + C_{\beta}^{en}}\right)^p \left(\frac{D/V_d}{C_{hs} + C_{\beta}^{en}} + 1\right)^q e^{-t}, p, q\right), \ t > 0$$
(11)

where

$$p = \frac{1}{k_{el}} \frac{C_{hs} + K_m}{C_{hs} + C_{\beta}^{en}}, \quad q = \frac{1}{k_{el}} \frac{C_{\beta}^{en} - K_m}{C_{hs} + C_{\beta}^{en}},$$

$$C_{\beta}^{en} = C_{hs} - \frac{r_{prod}}{k_{el}} + C_{\beta}.$$
(12)

**Proof** Since a single dose is added to the system, we have  $C(t) > C_{hs}$ , thus Eq. (8) can be transformed into

$$\left(\frac{p}{C(t) - C_{hs}} + \frac{q}{C(t) + C_{\beta}^{en}}\right) dC(t) = -dt,$$
(13)

with the notations p, q and  $C_{\beta}^{en}$  as defined in Eq. (12). It can be proved that p, q and  $C_{\beta}^{en}$  are positive, and  $C_{\beta}^{en} > K_m$  (see Appendix 2). Moreover,  $p + q = 1/k_{el}$  is the average time during which all drug compounds are assumed to be eliminated through the linear elimination pathway alone; p and q represent the partition of the time  $1/k_{el}$  modulated by  $\frac{C_{la}+K_m}{C_{la}+C_m^{en}}$  and  $\frac{C_{\beta}^m-K_m}{C_{la}+C_m^{en}}$ , respectively.

Integrating Eq. (13) from  $0^+$  to *t* leads to

$$p\ln(C(t) - C_{hs}) + q\ln(C(t) + C_{\beta}^{en})$$
  
=  $p\ln(C_0 - C_{hs}) + q\ln(C_0 + C_{\beta}^{en}) - t$  (14)

since  $C(t) > C_{hs}$ .

Equation (14) can be rearranged as

$$(C(t) - C_{hs})^{p} \left(C(t) + C_{\beta}^{en}\right)^{q} = (C_{0} - C_{hs})^{p} \left(C_{0} + C_{\beta}^{en}\right)^{q} e^{-t}$$
(15)

Dividing both sides of Eq. (15) by  $(C_{hs} + C_{\beta}^{en})^{p+q}$  gives rise to

$$\begin{aligned} &\left(\frac{C(t)-C_{hs}}{C_{hs}+C_{\beta}^{en}}\right)^{p} \left(\frac{C(t)-C_{hs}}{C_{hs}+C_{\beta}^{en}}+1\right)^{q} \\ &= \left(\frac{C_{0}-C_{hs}}{C_{hs}+C_{\beta}^{en}}\right)^{p} \left(\frac{C_{0}+C_{\beta}^{en}}{C_{hs}+C_{\beta}^{en}}\right)^{q} e^{-t}. \end{aligned}$$

In terms of X function, we have

$$\frac{C(t) - C_{hs}}{C_{hs} + C_{\beta}^{en}} = X\left(\left(\frac{C_0 - C_{hs}}{C_{hs} + C_{\beta}^{en}}\right)^p \left(\frac{C_0 + C_{\beta}^{en}}{C_{hs} + C_{\beta}^{en}}\right)^q e^{-t}, p, q\right), \quad t > 0$$
(16)

Substituting  $C_0$  with  $C_{hs} + D/V_d$  in Eq. (16), the closed form solution of C(t) shown in Eq. (11) is obtained.

#### Pharmacokinetic interpretation of $C_{\beta}^{en}$

 $C_{\beta}^{en}$  can be rewritten as

$$C_{\beta}^{en} = C_{hs} - C_{L,hs} + C_{\beta} \tag{17}$$

where  $C_{hs}$  is the baseline concentration given in Eq. (10);  $C_{L,hs} = r_{prod}/k_{el}$  is the baseline concentration of Eq. (8) if Michaelis–Menten kinetics is absent; and  $C_{\beta} = \frac{V_{max}}{k_{el}V_d} + K_m$ is a parameter defined for the model with no endogenous production in our previous study [19]. Specifically,  $C_{\beta}$  is the concentration at which a linear PK model with an elimination coefficient  $k_{el}$  can have the same rate of change as produced in another linear model at concentration  $K_m$ , which has an elimination rate constant  $k_{el} + \frac{V_{max}}{K_m V_d}$ .

In fact,  $C_{\beta}^{en}$  is the extension of  $C_{\beta}$  for the PK model that takes into account the endogenous production. Indeed, let us consider two linear models:

$$\frac{dC_1(t)}{dt} = k_{el}C_{hs} - k_{el}C_1(t) \tag{18}$$

and

$$\frac{dC_2(t)}{dt} = k_{el}C_{L,hs} - \left(k_{el} + \frac{V_{max}}{K_m V_d}\right)C_2(t).$$
(19)

Then,  $C_{\beta}^{en}$  is the concentration value of  $C_1(t)$  in Eq. (18) that gives the same change rate of concentration for  $C_2(t) = K_m$  in Eq. (19), i.e.,

$$\frac{dC_1(t)}{dt}\Big|_{C_1(t)=C_{\beta}^{en}} = \frac{dC_2(t)}{dt}\Big|_{C_2(t)=K_m}.$$
(20)

In short,  $C_{\beta}^{en}$  is the concentration of a linear model having endogenous production rate  $k_{el}C_{hs}$  and elimination coefficient  $k_{el}$ , that gives the same change rate of concentration for another linear model having endogenous production rate  $k_{el}C_{L,hs}$  and elimination coefficient  $k_{el} + \frac{V_{max}}{K_m V_d}$  at tion, we then have  $C_{hs} = C_{L,hs} = 0$ . In this case,  $C_{\beta}^{en} = C_{\beta}$ . We can also show that p and q generalize  $p_1$  and  $p_2$  given in [19], respectively.

### Area under the curve (AUC)

As there is a baseline concentration, the validity of a direct calculation of AUC from the observed concentrations using the trapezoidal rule has to be justified and adapted to this context. For drug compounds that are also endogenously produced, the use of the corrected concentrations has been recommended for the calculation of PK parameters. This corrected concentration,  $C(t) - C_{hs}$ , which is obtained by subtracting the baseline concentration from the observed concentration, is actually used to recover the exogenous drug contribution [25–27].

#### Partial area under the curve $(AUC_{0-t})$

We will first investigate the partial area under the curve from drug administration time zero until a certain time t, noted AUC<sub>0-t</sub>. Considering the current PK model where there is a baseline concentration, we can define AUC<sub>0-t</sub> as

$$AUC_{0-t} = \int_0^t (C(t) - C_{hs}) dt,$$
(21)

where C(t) is the observed concentration, and  $C_{hs}$  is the baseline concentration.

**Theorem 3** Given the PK model described by the Eqs. (8)–(9), the partial drug exposure AUC  $_{0-t}$  is

$$AUC_{0-t} = \frac{1}{k_{el}} \left( \frac{D}{V_d} + C_{hs} - C(t) \right) + \frac{C_{\beta}^{en} - K_m}{k_{el}} \ln \frac{C(t) + C_{\beta}^{en}}{D/V_d + C_{hs} + C_{\beta}^{en}},$$
(22)

where all parameters are as previously defined. Moreover, if Michaelis–Menten elimination pathway is absent, the corresponding partial area under the curve,  $AUC_{L,0-t}$ , becomes

$$AUC_{L,0-t} = \frac{D}{k_{el}V_d} \left(1 - e^{-k_{el}t}\right),$$
(23)

and we have

$$AUC_{0-t} < AUC_{L,0-t} \text{ if } V_{max} > 0, \text{ and } \lim_{V_{max} \to 0} AUC_{0-t} = AUC_{L,0-t}.$$
(24)

**Proof** The obtention of Eq. (22) is straightforward. We can multiply both sides of Eq. (13) by  $C(t) - C_{hs}$  and then make a simple rearrangement, which gives

$$(C(t) - C_{hs}) dt = -\left\{ p + q - q \frac{C_{hs} + C_{\beta}^{en}}{C(t) + C_{\beta}^{en}} \right\} dC(t).$$
(25)

Integration of Eq. (25) yields

$$\begin{aligned} AUC_{0-t} &= \int_{0}^{t} (C(t) - C_{hs}) \, dt \\ &= (p+q) \left( \frac{D}{V_d} + C_{hs} - C(t) \right) \\ &+ q(C_{hs} + C_{\beta}^{en}) \ln \frac{C(t) + C_{\beta}^{en}}{D/V_d + C_{hs} + C_{\beta}^{en}} \\ &= \frac{1}{k_{el}} \left( \frac{D}{V_d} + C_{hs} - C(t) \right) \\ &+ \frac{C_{\beta}^{en} - K_m}{k_{el}} \ln \frac{C(t) + C_{\beta}^{en}}{D/V_d + C_{hs} + C_{\beta}^{en}}. \end{aligned}$$

Furthermore, the model (Eqs. 8–9) satisfies

$$\frac{dC(t)}{dt} = r_{prod} - k_{el}C(t) - \frac{V_{max}C(t)}{V_d(K_m + C(t))} \le r_{prod} - k_{el}C(t),$$
(26)

with the same initial condition  $D/V_d + C_{hs}$ . This leads to the partial area under the curve of the linear model (Michaelis–Menten elimination is absent in Eq. (8)) as

$$AUC_{L,0-t} = \int_0^t \frac{D}{V_d} e^{-k_{el}t} dt = \frac{D}{k_{el}V_d} (1 - e^{-k_{el}t}).$$

By the Comparison Theorem [28], the solution of the current model (i.e. the concentration time curve) is upper bounded by that of the linear model, which gives rise to Eq. (24).  $\Box$ 

**Remark 2** AUC<sub>0-t</sub> given by Eq. (22) can be directly calculated using the *X* function, by applying the expression of C(t) given in Theorem 2.

#### Total drug exposure $(AUC_{0-\infty})$

The total drug exposure, as represented by  $AUC_{0-\infty}$  can be defined as

$$AUC_{0-\infty} = \int_0^\infty (C(t) - C_{hs}) dt, \qquad (27)$$

where C(t) and  $C_{hs}$  are the observed and baseline concentrations, respectively.

If Michaelis–Menten elimination pathway is absent in Eqs. (8)–(9), the following result is known for linear kinetics.

**Lemma 1** For the one-compartment PK model with linear elimination and constant endogenous production, the total

drug exposure,  $AUC_{L,0-\infty}$ , after a single IV bolus dose D, is

$$AUC_{L,0-\infty} = \frac{D}{k_{el}V_d}.$$
(28)

**Proof** Solving the linear PK model of

$$\frac{dC(t)}{dt} = r_{prod} - k_{el}C(t), \quad C(0^+) = D/V_d + C_{L,hs}$$
(29)

yields the corrected concentration as

$$C(t) - C_{L,hs} = \frac{D}{V_d} e^{-k_{el}t}, \quad t > 0.$$
 (30)

Accordingly, this total drug exposure is

$$AUC_{L,0-\infty} = \int_0^\infty (C(t) - C_{L,hs}) dt$$
  
= 
$$\int_0^\infty \frac{D}{V_d} e^{-k_e t} dt = \frac{D}{k_{el} V_d}.$$
 (31)

#### However, we have

**Theorem 4** For the considered PK model described by Eqs. (8)–(9), the total drug exposure over time defined by Eq. (27), AUC  $_{0-\infty}$ , is

$$AUC_{0-\infty} = \frac{D}{k_{el}V_d} - \frac{C_{\beta}^{en} - K_m}{k_{el}} \ln\left(1 + \frac{D/V_d}{C_{hs} + C_{\beta}^{en}}\right).$$
 (32)

Moreover, we have

$$AUC_{0-\infty} < AUC_{L,0-\infty} \text{ if } V_{max} > 0,$$
  
and 
$$\lim_{V_{max} \to 0} AUC_{0-\infty} = \frac{D}{k_{el}V_d} = AUC_{L,0-\infty}.$$
 (33)

**Proof** Integration of Eq. (25) from 0 to  $\infty$  yields

$$\begin{aligned} \operatorname{AUC}_{0-\infty} &= \int_0^\infty (C(t) - C_{hs}) \, dt \\ &= \int_0^\infty - \left\{ p + q - q \frac{C_{hs} + C_{\beta}^{en}}{C(t) + C_{\beta}^{en}} \right\} \, dC(t) \\ &= - (p + q)(C(\infty) - C_0) + q(C_{hs}) \\ &+ C_{\beta}^{en} \ln \left( \frac{C(\infty) + C_{\beta}^{en}}{C_0 + C_{\beta}^{en}} \right) \\ &= \frac{D}{k_{el}V_d} - \frac{C_{\beta}^{en} - K_m}{k_{el}} \ln \left( 1 + \frac{D/V_d}{C_{hs} + C_{\beta}^{en}} \right), \end{aligned}$$

where  $C(\infty) = C_{hs}$  and  $C_0 = C_{hs} + D/V_d$ .

Since  $C_{\beta}^{en} > K_m$ , the second term in the expression of  $AUC_{0-\infty}$  is positive when  $V_{max} > 0$ . Then we have

$$AUC_{0-\infty} < \frac{D}{k_{el}V_d} = AUC_{L,0-\infty}$$

Moreover, when the maximum velocity of Michaelis– Menten kinetics tends to zero, we have

$$C_{hs} \to \frac{r_{prod}}{k_{el}}$$
 and  $C_{\beta}^{en} \to K_m$ ,

which leads to

$$\lim_{V_{max}\to 0} \mathrm{AUC}_{0-\infty} = \frac{D}{k_{el}V_d} = \mathrm{AUC}_{L,0-\infty}.$$

**Remark 3** If the endogenous production rate  $r_{prod} = 0$ , then total drug exposure over time turns to

$$AUC_{0-\infty} = \frac{D}{k_{el}V_d} - \frac{C_{\beta} - K_m}{k_{el}} \ln\left(1 + \frac{D/V_d}{C_{\beta}}\right).$$
(34)

Moreover, if the endogenous production rate  $r_{prod}$  tends to infinity, we have

$$\lim_{r_{prod} \to \infty} AUC_{0-\infty} = \frac{D}{k_{el}V_d}.$$
(35)

**Remark 4** In fact,  $AUC_{0-t}$  can be rewritten as

$$AUC_{0-t} = \frac{D}{k_{el}V_d} - \frac{C_{\beta}^{en} - K_m}{k_{el}} \ln\left(1 + \frac{D/V_d}{C_{hs} + C_{\beta}^{en}}\right) - \frac{1}{k_{el}}(C(t) - C_{hs}),$$
(36)

which coincides with  $AUC_{0-\infty}$  in Eq. (32) when time tends to infinity.

# Impact of endogenous production on the corrected concentration and AUC

# Endogenous production and the corrected concentration

Though the use of the corrected concentration  $C(t) - C_{hs}$  seems a logic way for a fair estimation of exogenous compound's pharmacokinetics, its suitability and validation should be further investigated. The question would be to know if the corrected concentration is the same as the concentration generated by the system where no endogenous production is involved.

For this, two PK models are considered here: (1) the current PK model described by Eqs. (8)–(9); and (2) the linear model obtained by dropping the Michaelis–Menten elimination pathway from the former PK model. The following results are obtained.

- Linear pharmacokinetic model The corrected concentration is identical to the concentration generated by the system not involving endogenous production. It is understandable that for linear PK, endogenous production and exogenous administration contribute in parallel to the resulting drug concentration time course, a property known as the superposition principle for the linear time invariant system. Hence, the corrected concentration is a rational and fair choice for linear kinetics.
- Nonlinear pharmacokinetic model As observed in Fig. 1, the corrected concentration curves are different from the concentration curves generated by the systems with no endogenous production. Generally, these concentrations are even higher when  $r_{prod}$  increases (Fig. 1b). The ratio of the corrected concentration to that of the system with no endogenous production (Fig. 1d) grows exponentially with time for the system with endogenous production. As the superposition principle is no more valid for nonlinear PK, we can explain that a larger  $r_{prod}$  will make the accumulation of the resulting concentration even higher, and with exogenously administered drug fading over time, this becomes more imposing as reflected in the ratios. Therefore the corrected concentration is nonlinearly dependent on the endogenous production, even with a constant rate of endogenous production.

#### **Endogenous production and AUC**

For the PK model (Eqs. 8–9), we have found the explicit expressions of AUC<sub>0-t</sub> and AUC<sub>0- $\infty$ </sub> based on the corrected concentrations and established their relationships (Eqs. 24 and 32) with the corresponding linear PK model obtained by dropping the Michaelis-Menten elimination pathway. This relationship can be better perceived graphically. Indeed, AUC<sub> $0-\infty$ </sub> of the model (Eqs. 8–9) increases with  $r_{prod}$  in a sigmoid fashion but is always bounded above by  $AUC_{L,0-\infty}$ , which is the total drug exposure of the linear model (Fig. 2a). Moreover, the shapes of these AUC<sub>0- $\infty$ </sub> are controlled by  $V_{max}$  values. For a high  $V_{max}$ ,  $AUC_{0-\infty}$  is lower and needs a relatively high endogenous production  $r_{prod}$  to reach the saturate level given by  $AUC_{L,0-\infty}$ . Moreover it is obvious to see that  $AUC_{0-\infty}$ curve converges to the constant AUC<sub>L,0- $\infty$ </sub> when V<sub>max</sub> tends to zero, i.e., when the model tends to be linear as shown in Theorem 4.

It is interesting to explore how AUC<sub>0- $\infty$ </sub> responds to both endogenous production rate  $r_{prod}$  and exogenous dose *D*. For this, we calculated and displayed the contour plots of AUC<sub>0- $\infty$ </sub> vs.  $r_{prod}$  and *D* (Fig. 2b). For a large  $r_{prod}$ , the increase in AUC<sub>0- $\infty$ </sub> is almost equidistant for each increase

With Michaelis-Menten kinetics

(b)

log Conrrected Conc. (mIU/mI)

2

0

-2

-4

-6

5000

Ratio (no unit)

0

5



4000 3000 2000 1000 0 L 5 10 15 20 Time (h)

10

Time (h)

15

20

Fig. 1 (Online color) The impact of  $r_{prod}$  on the corrected concentration for linear and nonlinear PK models for  $r_{prod} = 0, 15, 30 \text{ mIU}/$ ml/h. D = 1350 mIU/kg, $V_d = 61.18 \text{ ml/kg};$  $k_{el} = 0.21/h,$  $V_{max} = 1993$  mIU/h/kg and  $K_m = 67.23$  mIU/ml. **a** Time courses



Contours of  $AUC_{0-\infty}$ (b) 1800 150 8 1600 1400

of log corrected concentration for the linear models; b time courses of

log corrected concentration for the nonlinear models; c ratios for the

linear models; d ratios for the nonlinear models



Fig. 2 (Online color) Drug exposure versus endogenous production rate  $r_{prod}$  and administered dose D. a AUC<sub>0- $\infty$ </sub> versus  $r_{prod}$  for  $V_{max} = 0,2000, 6000 \text{ mIU/h/kg}, D = 1375 \text{ mIU/kg}; \mathbf{b}$  contour plot

of AUC<sub>0- $\infty$ </sub> in function of  $r_{prod}$  and D, where  $V_{max} = 2000$  mIU/h/kg. The other parameters are the same as those in Fig. 1



**Fig. 3** (Online color) Partial area under the curve  $AUC_{0-t}$  versus time t or endogenous production rate  $r_{prod}$ . **a**  $AUC_{0-t}$  versus time t for different  $r_{prod}$  as 0, 10, 20, 30 mIU/ml/h; **b**  $AUC_{0-t}$  versus endogenous production rate  $r_{prod}$  for time t = 1, 4, 12, 48 h. Except time t

in D, hence proportional to the dose. However, this linear increase is not maintained for low levels of  $r_{prod}$ . This phenomenon can be explained with the concept of dominant elimination pathway that we found in [19], where the linear and nonlinear elimination pathways can have their dominant role altered depending on drug concentration levels. A large value of  $r_{prod}$  leads to a high level of plasma concentration, making thus the first-order elimination dominant. However, when the clearance from the linear elimination pathway  $(k_{el}V_d)$  is less than the intrinsic clearance following Michaelis–Menten kinetics  $(V_{max}/K_m)$ , a small  $r_{prod}$  leads to a low baseline concentration, thus giving rise to a low level of drug plasma concentration where Michaelis-Menten elimination pathway dominates, and this is particularly true for a small dose D. Moreover, for a given AUC<sub>0- $\infty$ </sub>, dose can be estimated as D = $k_{el}V_d \cdot AUC_{0-\infty}$  when  $r_{prod} \rightarrow \infty$ , while dose D can only be numerically estimated from Eq. (34) when  $r_{prod} = 0$ .

Similar observations can be found and explained for the partial  $AUC_{0-t}$  (Fig. 3).

#### The net total drug exposure (AUC<sub>net</sub>)

We have calculated the baseline concentration  $C_{hs}$  for the steady state of Eq. (8). At this state, the system elimination is balanced with the endogenous input rate  $r_{prod}$ . Thus the validity of using  $C_{hs}$  as a proxy of the contribution of endogenous input can only be established when the system is not altered by any exogenous drug input. For a constant  $r_{prod}$ ,  $C_{hs}$  stays constant. However, in the presence of exogenous drug administration, the fair share of endogenous input in the whole drug exposure can not be constant.





and endogenous production rate  $r_{prod}$ , other parameters are fixed as D = 1375 mIU/kg,  $k_{el} = 0.21 \text{ /h}$ ,  $V_{max} = 1993 \text{ mIU/h/kg}$ ,  $K_m = 67.23 \text{ mIU/ml}$ ,  $V_d = 61.18 \text{ ml/kg}$  [9]

Instead, the transient endogenous contribution should be a function of time that varies with the PK of the exogenous drug input. It is hence important to differentiate the fair contributions of endogenous and exogenous inputs to the total drug exposure.

Estimation of the contribution of endogenous production to the total drug exposure cannot be directly obtained as the case for the determination of the baseline concentration, where only several blood samples collected prior to drug administration are needed. However, in the actual modeling framework, we can separately model the contribution of endogenous and exogenous inputs, and numerically calculate the net drug exposure from exogenous contribution by removing the endogenous part.

In fact, the current model (Eqs. 8–9) can be further separated into the following submodel

$$\frac{d}{dt}C_{en}(t) = r_{prod} - k_{el}C_{en}(t) - \frac{V_{max}C_{en}(t)}{V_d(K_m + C_{en}(t) + C_{ex}(t))},$$
  

$$C_{en}(0) = C_{hs},$$

$$\frac{d}{dt}C_{ex}(t) = -k_{el}C_{ex}(t) - \frac{V_{max}C_{ex}(t)}{V_d(K_m + C_{en}(t) + C_{ex}(t))},$$

$$C_{ex}(0^+) = D/V_d,$$
(38)

which describe the concentrations  $C_{en}(t)$  and  $C_{ex}(t)$ , generated by endogenous and exogenous inputs, respectively. Note that adding Eqs. (37) and (38) gives the original model, where  $C = C_{en} + C_{ex}$ . Moreover, the non-

(37)



Fig. 4 (Online color) Profiles of time course of observed, endogenous and exogenous concentrations and impact of endogenous production on AUC. **a** Time course of observed, endogenous and exogenous concentrations in the case of  $r_{prod} = 50$  mIU/ml/h simulated from

negativity of the submodel and local stability of the unique steady state  $E^* = (C_{en}^*, C_{ex}^*) = (C_{hs}, 0)$  are given in Appendix 3.

Then the net total drug exposure should be

$$AUC_{net} = \int_0^\infty (C(t) - C_{en}(t)) \, dt = \int_0^\infty C_{ex}(t) \, dt.$$
(39)

Based on Eqs. (37)–(38),  $C_{en}(t)$  and  $C_{ex}(t)$  are simulated numerically to estimate the net drug exposure, and the results are displayed in Fig. 4.

In Fig. 4a, we can observe the influence of exogenous drug input on the PK of endogenous production. Instead of being an independent concentration curve of constant value  $C_{hs}$ ,  $C_{en}(t)$  is driven by the PK of exogenous drug administration. In fact, initially at the value of  $C_{hs}$ ,  $C_{en}(t)$  increases after the dose administration and follows a bell curve then decreases towards  $C_{hs}$ . This justifies our argument on the improper use of  $C_{hs}$  in the estimation of drug exposure under the conditions of the current model. In Fig. 4b,  $AUC_{0-\infty}$ estimated using the baseline concentration  $C_{hs}$  and AUC<sub>net</sub> are reported versus  $r_{prod}$ . It clearly shows that AUC<sub>0- $\infty$ </sub> overestimates the drug exposure of the exogenous administration. However, it is still interesting to see that, even when the endogenous contribution is more properly dropped, AUC<sub>net</sub> remains dependent on  $r_{prod}$ , which is in fact increasing to finally saturate with  $r_{prod}$ . The nonlinear



the model (Eqs. 37–38). **b** Impact of endogenous production  $(r_{prod} \in [0,300 \text{ mIU/ml/h}])$  on AUC<sub>0- $\infty$ </sub> and AUC<sub>net</sub>. In all simulations, other parameters are fixed as D = 1375 mIU/ml,  $k_{el} = 0.21$ /h,  $V_{max} = 2000 \text{ mIU/h/kg}$ ,  $K_m = 67.23 \text{ mIU/ml}$ ,  $V_d = 61.18 \text{ ml/kg}$ 

Michaelis–Menten elimination pathway, which saturates at high concentration values, is responsible for this unusual phenomenon. With a large  $r_{prod}$ , drug concentration is inclined to accumulate to reach a high level such that the drug is less easily eliminated, leading to a higher AUC<sub>net</sub>. Moreover, for a very large  $r_{prod}$ , linear pathway dominates the whole drug elimination. In this case, the system is likely to manifest a linear PK and reaches a plateau of AUC<sub>net</sub>.

### **Discussion and conclusions**

In the current paper, we have formulated the closed form solution for the one-compartment PK model involving simultaneous first-order and Michaelis–Menten elimination with endogenous production in the case of a single IV dose. This model extends the one that we have previously studied with the additional consideration here of the endogenous production [19]. The extension is physiologically natural since it allows to mathematically outreach to those substances that are also endogenously produced. While our progress in this direction is a significant step forward, this is not the end of the story since more refined drug models and modeling approaches are being proposed and for which a rigorous mathematical analysis is still lacking [3, 29–32]. Such models could include multiple compartments [32],

involving feedback mechanisms regulating endogenous production [3, 33–35], or target-mediated drug disposition (TMDD) [29–31]. Though endogenous production is considered constant here, this work definitely provides a foundation for the understanding of the influence of endogenous production on PK and sheds light on the inner structure of PK components and their intertwined relationships.

Our work indicates the need to revisit and update the estimation of relevant PK parameters, especially for the case of more complex drugs exhibiting non-linear kinetics. In the presence of endogenous production, the current study shows that it can have a great impact on the estimation of drug exposure, whether using the corrected concentration or net exogenous concentration curve. A further practical strategy to rationally estimate drug exposure needs to be developed.

As previously mentioned, the introduction of the X function was motivated by the usefulness of the Lambert W function to express the closed form solution of onecompartment PK models involving the Michaelis-Menten elimination alone [17]. In fact, through a limiting process, we were able to show that the X function can be linked to the Lambert W function. We proved that this newly introduced X function was also suitable for the expression of the closed form solution of one-compartment PK models with simultaneous first-order and Michaelis-Menten elimination [19]. In the current paper, we have shown that the use of X function can be further extended to the case of endogenous production. Indeed, the X function has its own specific mathematical properties such as the real branches as illustrated in the Appendix 4 using typical parameter values. More complex branches can be explored as what is known for the case of the Lambert W function [18]. For numerical purposes, we have implemented the X function into Matlab, with the aim to make it accessible to the users.

**Acknowledgements** This work is supported by FRQNT Fellowship (X. Wu) and the NSERC-Industrial Chair in Pharmacometrics (cofunded by InVentiv Health and Pfizer) and FRQNT Projet d'équipe led by F. Nekka. X. Wu and J. Li also thank the support from NSFC (No.11501358). We thank the referees for their careful reading and valuable comments which helped improving the quality of the paper.

# Appendix 1: Well-posedness of the model (Eq. 8)

**Lemma 2** For the model (Eq. 8), we have the following results:

- (i) There is a unique positive solution  $C_{hs}$  as shown in Eq. (10).
- (ii) C(t) > 0 for all t > 0 provided that C(0) > 0 and  $\lim_{t\to\infty} C(t) = C_{hs}$ .

703

letting the right hand be zero yields 1 - V = C(x)

$$r_{prod} - k_{el}C(t) - \frac{1}{V_d} \frac{V_{max}C(t)}{K_m + C(t)} = 0$$

This can be further transformed into the following quadratic equation

$$C^{2}(t) + \left(\frac{V_{max}}{k_{el}V_{d}} + K_{m} - \frac{r_{prod}}{k_{el}}\right)C(t) - \frac{r_{prodK_{m}}}{k_{el}} = 0.$$

Since the product of the roots of the above quadratic equation is  $-\frac{r_{prodK_m}}{k_{el}} < 0$ , hence the roots must be real with one negative and one positive  $C_{hs}$  as shown in Eq. (10).

(ii) If  $C(0) = C_{hs} > 0$ , then  $\frac{dC(t)}{dt}|_{t=0} = 0$  implying  $C(t) = C_{hs} > 0$  for all t > 0. If  $C(0) > C_{hs} > 0$ , we have  $\frac{dC(t)}{dt}|_{t=0} < 0$  implying a decrease of C(t) at t = 0. By the continuity of solution, C(t) will continue to decrease as long as  $C(t) > C_{hs}$  for t > 0, and will eventually converge to  $C_{hs}$  since  $\frac{dC(t)}{dt}|_{C(t)=C_{hs}} = 0$ . Therefore, in this case C(t) is a strictly decreasing sequence and has  $C_{hs}$  as a lower bound. By the monotone convergence theorem, we have  $\lim_{t\to\infty} C(t) = C_{hs}$ . Similarly, if  $0 < C(0) < C_{hs}$ , C(t) increases and eventually converges to  $C_{hs}$ .

# Appendix 2: Claim "p, q and $C_{\beta}^{en}$ are positive and $C_{\beta}^{en} > K_m$ "

**Proof**  $C_{L,hs}$  satisfies  $r_{prod} = k_{el}C_{L,hs}$ , and  $C_{hs}$  satisfies  $r_{prod} = k_{el}C_{hs} + \frac{V_{max}}{V_d} \frac{C_{hs}}{K_m + C_{hs}}$ , thus we have  $k_{el}(C_{hs} - C_{L,hs}) = -\frac{V_{max}}{V_d} \frac{C_{hs}}{K_m + C_{hs}}$ .

This yields

$$C_{hs}-C_{L,hs}=-rac{V_{max}}{k_{el}V_d}rac{C_{hs}}{K_m+C_{hs}}.$$

Accordingly, we have

$$\begin{split} C^{en}_{\beta}-K_m &= C_{hs}-C_{L,hs}+C_{\beta}-K_m \\ &= -\frac{V_{max}}{k_{el}V_d}\frac{C_{hs}}{K_m+C_{hs}} + \frac{V_{max}}{k_{el}V_d} = \frac{V_{max}}{k_{el}V_d}\frac{K_m}{K_m+C_{hs}} > 0. \end{split}$$

It is thus obvious that p and q are positive.

# Appendix 3: Stability of the steady state solution of the model (Eqs. 37–38)

**Lemma 3** For the model (Eqs. 37–38), the solution is nonnegative for any initial conditions  $C_{en}(0) = C_{hs} > 0$  and



**Fig. 5** Real branches of *Lambert W* and *X* functions. Left panel: *Lambert W* function has two real branches,  $W_0 \in (-1, \infty)$  for  $z \in (-1/e, \infty)$  and  $W_{-1} \in (-\infty, -1)$  for  $z \in (-1/e, 0)$ . Right panel: *X* function (p = 2/3, q = 1/3) has three real branches,  $X_0 \in (0, \infty)$  for

 $C_{ex}(0) > 0$ , and there is a unique equilibrium  $E^* = (C^*_{en}, C^*_{ex}) = (C_{hs}, 0)$  which is locally asymptotically stable.

**Proof** The nonnegativity of  $C_{en}(t)$  and  $C_{ex}(t)$  follows immediately from Theorem 5.2.1 on page 81 in [28]. Letting the right hands of Eqs. (37)–(38) be zeroes, we can simply have  $C_{en}^* = C_{hs}$  and  $C_{ex}^* = 0$ .

The linearized the system at the equilibrium  $E^*$  is

$$\begin{cases} C'_{en}(t) &= -\left(k_{el} + \frac{V_{max}}{V_d} \frac{K_m}{(K_m + C_{hs})^2}\right) C_{en}(t) + \frac{V_{max}}{V_d} \frac{C_{hs}}{(K_m + C_{hs})^2} C_{ex}(t), \\ C'_{ex}(t) &= -\left(k_{el} + \frac{V_{max}}{V_d} \frac{1}{K_m + C_{hs}}\right) C_{ex}(t). \end{cases}$$

Thus, the Jacobian matrix is

$$J(E^*) = \begin{pmatrix} -(k_{el} + \frac{V_{max}}{V_d} \frac{K_m}{(K_m + C_{hs})^2}) & \frac{V_{max}}{V_d} \frac{C_{hs}}{(K_m + C_{hs})^2} \\ 0 & -(k_{el} + \frac{V_{max}}{V_d} \frac{1}{K_m + C_{hs}}) \end{pmatrix}$$

and the two eigenvalues  $\lambda_1$  and  $\lambda_2$  satisfy

$$\lambda_1 + \lambda_2 = tr(J(E^*)) < 0$$
 and  $\lambda_1 \lambda_2 = det(J(E^*)) > 0.$ 

These imply  $\lambda_1$  and  $\lambda_2$  are negative, which ensures the local asymptotic stability of  $E^*$ .

# Appendix 4: Real branches of *Lambert W* and *X* functions

*Lambert W* function is known to have two real branches, with one branch  $W_0 \in (-1, \infty)$  for *z* defined in  $(-1/e, \infty)$ , and the other  $W_{-1} \in (-\infty, -1)$  for *z* defined in (-1/e, 0) (left panel, Fig. 5) [17, 18]. However, the case of



 $z \in (0,\infty), X_{-1} \in (-2/3,0)$  for  $z \in (0, 1/3 \times 4^{1/3})$  and  $X_{-2} \in (-\infty, -2/3)$  for  $z \in (-\infty, 1/3 \times 4^{1/3})$ 

X function is more complex. Without going into details, we simply illustrate here a case of real branches of an X function, with the aim to show its particularity compared to *Lambert W* function (right panel, Fig. 5).

As observed in the right panel of Fig. 5, there is only one real branch in the first quadrant. In fact, this is true for all X functions. Taking the derivative of X function, we have

$$\frac{d}{dz}X(z,p,q) = \frac{1}{z} \cdot \left(\frac{p}{X(z,p,q)} + \frac{q}{1 + X(z,p,q)}\right)^{-1} > 0,$$
(40)

which guarantees there is only one real branch in the first quadrant. In the current study, we are interested only in this real branch  $X(z, p, q) \in (0, \infty)$  and use it to express the analytical solutions of PK models. Thus we use the letter *X* to denote this unique real branch in the current paper.

#### References

- Derendorf H, Meibohm B (1999) Modeling of pharmacokinetic/ pharmacodynamic (PK/PD) relationships: concepts and perspectives. Pharm Res 16(2):176–185
- Li J, Nekka F (2013) A rational quantitative approach to determine the best dosing regimen for a target therapeutic effect: a unified formalism for antibiotic evaluation. J Theor Biol 319:88–95
- Craig M, Humphries AR, Nekka F, Bélair J, Li J, Mackey MC (2015) Neutrophil dynamics during concurrent chemotherapy and G-CSF administration: mathematical modelling guides dose optimisation to minimise neutropenia. J Theor Biol 385:77–89
- Câmara De Souza D, Craig M, Cassidy T, Li J, Nekka F, Bélair J, Humphries AR (2018) Transit and lifespan in neutrophil

production: implications for drug intervention. J Pharmacokinet Pharmacodyn 45(1):59–77

- Krzyzanski W, Wiczling P, Lowe P, Pigeolet E, Fink M, Berghout A, Balser S (2010) Population modeling of filgrastim PK-PD in healthy adults following intravenous and subcutaneous administrations. J Clin Pharmacol 50:101S–112S
- Scholz M, Schirm S, Wetzler M, Engel C, Loeffler M (2012) Pharmacokinetic and -dynamic modelling of G-CSF derivatives in humans. Theor Biol Med Model 9:32
- Perreault S, Burzynski J (2009) Romiplostim: a novel thrombopoiesis-stimulating agent. Am J Health Syst Pharm 66:817–824
- Platanias LC, Miller CB, Mick R, Hart RD, Ozer H, McEvilly JM, Jones RJ, Ratain MJ (1991) Treatment of chemotherapyinduced anemia with recombinant human erythropoietin in cancer patients. J Clin Oncol 9(11):2021–2026
- Woo S, Krzyzanski W, Jusko WJ (2006) Pharmacokinetic and pharmacodynamic modeling of recombinant human erythropoietin after intravenous and subcutaneous administration in rats. J Pharmacol Exp Ther 319(3):1297–1306
- Gouyette A, Kerr DJ, Kaye SB, Setanoians A, Cassidy J, Bradley C, Forrest G, Soukop M (1988) Flavone acetic acid: a nonlinear pharmacokinetic model. Cancer Chemother Pharmacol 22(2):114–119
- Lee BY, Kwon KI, Kim MS, Baek IH (2016) Michaelis-Menten elimination kinetics of etanercept, rheumatoid arthritis biologics, after intravenous and subcutaneous administration in rats. Eur J Drug Metab Pharmacokinet 41:433–439
- Wagner JG, Gyves JN, Stetson PL, Walker-Andrews SC, Wollner IS, Cochran MK, Ensminger WD (1986) Steady-state nonlinear pharmacokinetics of 5-fluorouracil during hepatic arterial and intravenous infusion in cancer patients. Cancer Res 46:1499–1506
- Valodia PN, Seymour MA, McFadyen ML, Miller R, Folb PI (2000) Validation of population pharmacokinetic parameters of phenytoin using the parallel Michaelis–Menten and first-order elimination model. Ther Drug Monit 22(3):313–319
- Beal SL (1982) On the solution to the Michaelis–Menten equation. J Pharmacokin Biopharm 10:109–119
- Beal SL (1983) Computation of the explicit solution to the Michaelis–Menten equation. J Pharmacokin Biopharm 11:641–657
- Schnell S, Mendoza C (1997) Closed form solution for timedependent enzyme kinetics. J Theor Biol 187:207–212
- Tang S, Xiao Y (2007) One-compartment model with Michaelis– Menten elimination kinetics and therapeutic window: an analytical approach. J Pharmacokinet Pharmacodyn 34:807–827
- Corless RM, Gonnet GH, Hare DEG, Jeffrey DJ, Knuth DE (1996) On the Lambert W function. Adv Comput Math 5:329–359
- Wu X, Li J, Nekka F (2015) Closed form solutions and dominant elimination pathways of simultaneous first-order and Michaelis– Menten kinetics. J Pharmacokinet Pharmacodyn 42:151–161
- Foley C, Mackey MC (2009) Mathematical model for G-CSF administration after chemotherapy. J Theor Biol 257:27–44

- Wright EM (1949) The linear difference-differential equation with constant coefficients. Proc R Soc Edinburgh A62:387–393
- Asl F, Ulsoy AG (2003) Analysis of a system of linear delay differential equations. J Dyn Syst Meas Control 125(2):215–223
- Dostalek M, Gardner I, Gurbaxani BM, Rose RH, Chetty M (2013) Pharmacokinetics, pharmacodynamics and physiologically-based pharmacokinetic modelling of monoclonal antibodies. Clin Pharmacokinet 52(2):83–124
- Kozawa S, Yukawa N, Liu J, Shimamoto A, Kakizaki E, Fujimiya T (2007) Effect of chronic ethanol administration on disposition of ethanol and its metabolites in rat. Alcohol 41(2):87–93
- European Medicines Agency (2010) http://www.ema.europa. eu/docs/en\_GB/document\_library/Scientific\_guideline/2010/01/ WC500070039.pdf
- 26. FDA Guidance: Guidance for Industry. Bioavailability and Bioequivalence Studies Submitted in NDAs or INDs-General Considerations (2014) http://www.fda.gov/downloads/drugs/gui dancecomplianceregulatoryinformation/guidances/ucm389370. pdf
- Health Canada: Guidance Document: Conduct and Analysis of Comparative Bioavailability Studies (2012) http://www.hc-sc.gc. ca/dhp-mps/alt\_formats/pdf/prodpharma/applic-demande/guide-ld/ bio/gd\_standards\_ld\_normes-eng.pdf
- Smith HL (1995) Monotone dynamical systems: an introduction to the theory of competitive and cooperative systems, vol 41. Mathematical surveys and monographs. AMS, Providence
- Aston PJ, Derks G, Agoram BM, van der Graaf PH (2014) A mathematical analysis of rebound in a target-mediated drug disposition model: I. Without feedback. J Math Biol 68(6):1453–1478
- Patsatzis DG, Maris DT, Goussis DA (2016) Asymptotic analysis of a target-mediated drug disposition model: algorithmic and traditional approaches. Bull Math Biol 78(6):1121–1161
- Peletier LA, Benson N, van der Graaf PH (2009) Impact of plasma-protein binding on receptor occupancy: an analytical description. J Theor Biol 256:253–262
- Wu X, Nekka F, Li J (2016) Steady-state volume of distribution of two-compartment models with simultaneous linear and saturated elimination. J Pharmacokinet Pharmacodyn 43(4):447–459
- 33. Quartino AL, Karlsson MO, Lindman H, Friberg LE (2014) Characterization of endogenous G-CSF and the inverse correlation to chemotherapy-induced neutropenia in patients with breast cancer using population modeling. Pharm Res 31(12):3390–3403
- Hareng L, Hartung T (2002) Induction and regulation of endogenous granulocyte colony-stimulating factor formation. Biol Chem 383(10):1501–1517
- Roberts AW (2005) G-CSF: a key regulator of neutrophil production, but that's not all!. Growth Factors Chur Switz 23(1):33–41